



Plant Archives

Journal homepage: <http://www.plantarchives.org>

DOI Url : <https://doi.org/10.51470/PLANTARCHIVES.2025.v25.no.1.172>

EFFECT OF PHYTO-EXTRACTS AND BIO-AGENTS ON BIOCHEMICALS, ROOT ROT INCIDENCE AND *MELOIDOGYNE* POPULATION OF FENNEL (*FOENICULUM VULGARE* MILL.)

Kiran Kumawat^{1*}, S.K. Goyal¹, B.S. Chandrawat², G.K. Mittal³, R.P. Ghasolia¹, Lalita Lakhran⁴,
B.L. Naga⁵, Dinesh Kumar Meena¹ and Sushila Yadav⁶

¹Department of Plant Pathology, SKN College of Agriculture, S.K.N.A.U., Jobner, Rajasthan, India.

²Department of Nematology, SKN College of Agriculture, S.K.N.A.U., Jobner, Rajasthan, India.

³Department of Biochemistry, SKN College of Agriculture, SKNAU, Jobner, Rajasthan, India.

⁴Department of Plant Pathology, Agricultural Research Station, Mandor, A.U., Jodhpur, Rajasthan, India.

⁵Department of Entomology, SKN College of Agriculture, SKNAU, Jobner, Rajasthan, India.

⁶Department of Plant Pathology, Rajasthan College of Agriculture, MPUAT, Udaipur, Rajasthan, India.

*Corresponding author E-mail : kkumawatkiran666@gmail.com

(Date of Receiving-05-12-2024; Date of Acceptance-18-02-2025)

ABSTRACT

Seed spices are very important for human health and widely grown throughout the country. Among these, fennel (*Foeniculum vulgare* Mill.) is one of the also important seed spices belong to the *Apiaceae* family. Root rot is the most common disease of fennel caused by *Fusarium solani* and its severity is aggravated by the presence of root rot nematode. In lieu of this, an experiment was conducted to manage *F. solani* in association with *Meloidogyne javanica* through bio-agents and phyto-extracts. The bio-agents (*Pseudomonas fluorescens* and *Trichoderma harzianum*) were applied through soil (2.5 kg/ha), while botanicals (garlic clove and parthenium leaves extract) through seed soaking (@10% for 30 min.). Maximum elevated growth parameters of fennel were recorded with *T. harzianum* + garlic extract followed by parthenium extract. The lowest root rot incidence (11.48%), reduced nematode population (163.47 eggs) and enhanced the level of PO (0.788 $\mu\text{mol}/\text{min}/\text{g}$), PPO (0.094 $\mu\text{mol}/\text{min}/\text{g}$), PAL (0.646 $\mu\text{mol}/\text{min}/\text{g}$) and phenol (3.110 mg/g) contents were recorded in *T. harzianum* + garlic extract treated roots of fennel plants over untreated check (0.423, 0.027, 0.117 $\mu\text{mol}/\text{min}/\text{g}$ and 1.513 mg/g, respectively) followed by *T. harzianum* + parthenium extract. In conclusion, the use of the *T. harzianum* + garlic extract can be considered as a tool complementary for the commercial management of root rot of fennel in areas where this disease is gaining importance and becoming a limiting factor. Additionally, it can be used for managing pathogen resistance as well as also provide opportunity for organic production of spices for consumers.

Key words : Fennel, Root rot, *Fusarium solani*, *Meloidogyne javanica*, Bio-agents, Phyto-extracts.

Introduction

Among the seed spices, fennel (*Foeniculum vulgare* Mill.) is one of the important crops belongs to the *Apiaceae* family and originated from the Mediterranean region (Omidbaigi, 2000). In India, area under fennel cultivation is around 83 thousand hectares with a production of 137 thousand metric tonnes and productivity of 1.7 metric tonnes/ha, while in Rajasthan, area is around 30,814 hectares with 33,563 tonnes production and 1089

kg/ha productivity (Anonymous, 2020-21). During all the growth stages of fennel crop, right from germination to maturity is attacked by a large spectrum of diseases caused by fungi, bacteria, viruses and nematodes (Khare *et al.*, 2014). Among diseases, root rot caused by *Fusarium solani* is a major fungal disease. The incidence of root rot was reported 20 to 25 per cent in fennel (Gupta *et al.*, 1976). Root knot nematode, *Meloidogyne* spp. was reported on fennel and cumin crop (Patel, 2011; Kant

et al., 2013). Under tropical and subtropical conditions root knot nematode causes much higher damage (Taylor *et al.*, 1978). In India, 21.07 per cent average loss caused by root knot nematode on fennel (Patel *et al.*, 2002). There are several reports of complex disease development with nematode and fungus interaction in various crops *i.e.*, tomato (Bhagawati *et al.*, 2000; Meena *et al.*, 2021a), pea (Akhtar *et al.*, 2007), black gram (Kumar *et al.*, 2017), fennel (Kumawat *et al.*, 2022) and cucumber (Meena *et al.*, 2023). During early phase of research, Atkinson (1892) stated that disease incidence of root rot may be increased by simultaneous or sequential infection by root knot nematodes and *Fusarium* spp. Later on, it was noted that nematodes facilitate fungus to invade plant roots and provoked disease severity by plant roots modification (Mai *et al.*, 1987). Avoiding crop losses due to diseases caused by pathogenic fungi, bacteria, viruses, viroids and nematodes is one of the most significant issues in getting economic production of plant. At present, fast and effective control of fungal and other diseases in the crops is usually achieved using synthetic pesticides. Thus, chemical control of fungal diseases is largely dependent on the use of systemic and non-systemic fungicides. Furthermore, chemicals of this type evoke undesirable effects on the environment and leave residues, toxic to mammals in the products and risk of developing resistance in microorganisms.

Many organic amendments, plant extracts and bio-agents or their combination are the most effective against the nematode fungus complex diseases in different crops. Soil population pathogen and root galling were significantly suppressed by application of bio-agents and botanicals. Therefore, some of the phyto-extracts and bioagents were tried to manage the disease considering environmentally safe and eco-friendly.

Materials and Methods

Botanicals

The leaves of Parthenium and garlic cloves were collected, washed water and grind in absolute alcohol in equal volume. The grinded material was kept at room temperature for 48 h and filtered through filter paper. The solvent, absolute alcohol was evaporated at 70°C from the extract so as to get a semi-solid material. This semi-solid material was considered as stock solution for further experimentation with required concentration.

Culture of *Fusarium solani*

The root rot causing fungus (*Fusarium solani*) was isolated from diseased roots of fennel and further purified and identified on the basis of cultural and morphological characters. For mass multiplication of fungus, sorghum

grains were boiled partially and then spread on clean blotting paper for drying. About 250g of such sorghum grains were filled in each 1000ml flask and autoclaved at 15 psi pressure for 30 minutes. The mycelial bit (5 mm dia.) of 7 days old culture of *Fusarium solani* was inoculated under aseptic conditions on grains and incubated at 25±1°C for 7 days. Meanwhile the flasks were jiggled to avoid clumping of grains and to facilitates early growth of fungus.

Culture of *Meloidogyne javanica*

The fennel plants infected with root-knot nematode were collected from the field and brought to the laboratory. The infected roots were drenched in water and egg masses were carefully detached from roots. Later, infected roots having female nematodes were stained in 0.1 per cent acid fuchsin lactophenol and destained in pure lactophenol (McBeth *et al.*, 1941). Perineal patterns of adult females were cut using a scalpel and removed the body contents using camel hair brush No. 1 (Taylor *et al.*, 1974) and prepared a mount on glass slides. Observations of the perineal patterns showed the presence of *M. javanica* species. Simultaneously, collected egg masses were put into watch glasses in distilled water at laboratory temperature for hatching. Newly hatched 2nd stage juveniles (J₂) were then inoculated on already grown and maintained one month old fennel plants in pots filled with sterilized soil to acquired *M. javanica* pure population to carry out further experiments.

Biochemical analysis

The determination of peroxidase (EC 1.11.1.7) activity (Hammerschmidt *et al.*, 1982), polyphenol oxidase (EC 1.10.3.1) (Mayer *et al.*, 1965), phenylalanine ammonia lyase (PAL) (EC 4.3.1.5) (Dickerson *et al.*, 1984) and phenol activity (Malik and Singh, 1980) were estimated by following standard protocols.

Bio-agents and botanicals in managing root rot disease and root-knot nematode

In this experiment, two bio-agents (*Pseudomonas fluorescens* and *Trichoderma harzianum*) and two phyto-extracts (garlic clove and parthenium leaves) were used alone as well as in combination. The bioagents were applied through soil (2.5 kg/ha) while botanicals through seed soaking (@ 10 % for 30 min.). Plants were uprooted sixty days after sowing and observations on growth characters, per cent root rot incidence, nematode reproduction and on accumulation of peroxidase, polyphenol oxidase, phenylalanine ammonia lyase and phenol were estimated in fennel roots using spectrophotometer.

Statistical analysis

Data were statistically analyzed for interpretation of findings. The critical difference was calculated for comparison of treatments at 5% level of significance.

Results and Discussion

The data of the investigation indicated that all bio-agents (2.5 kg/ha in soil) and botanicals (seed soaking in 10 % for 30 min), applied alone or in combination, significantly increased plant growth parameters and decreased root rot incidence and nematode reproduction as compared to check (Table 1). Among these, soil application of *T. harzianum* (2.5 kg/ha) + seed soaking in garlic extract (10 % for 30 min) was recorded highly effective in providing maximum shoot length (25.50 cm), shoot weight (3.51 g), root length (21.94 cm), root weight (2.35 g) along with minimum root rot incidence (11.48 %), number of nematode galls/plant (6.67), number of egg masses/plant (3.67), number of eggs/egg mass (238.67) and number of juveniles/200 cc soil as compared to check (6.24cm, 0.43g, 4.70cm, 0.37g, 71.11%, 29.34, 23.34, 298.67 and 874.34, respectively). The present findings are in accordance with the results of earlier researchers (Kamal *et al.*, 2009; Park *et al.*, 2014; Meena *et al.*, 2015; Arya, 2016; Baheti *et al.*, 2016; Kumhar *et al.*, 2018a & b; Bhati *et al.*, 2022a & b). Kamal *et al.* (2009) observed effectiveness of neem oil in inhibiting root-knot nematode population in tomato while Park *et al.* (2014) suggested that *M. hapla* of tomato and carrot could be managed by application of biocontrol agent (*Bacillus cereus*). Similarly, reduced the infection of *M. incognita* and germination percentage in tomato increased with the combined application of *Trichoderma harzianum* and vermicompost (Arya, 2016). Use of plant extracts not only increase the maize growth but also decrease the nematode infection. Meena *et al.* (2015) applied neem leaves powders (4 q/ha) and found to be the most effective in improving growth of maize and reducing infection of *Heterodera zaeae*. Baheti *et al.* (2016) observed neem leaves powder (@ 20 per cent w/w) as most effective in increasing maize growth and in reducing infection of cyst nematode in maize. Meena *et al.* (2016) have also reported that application of carbosulfan (@ 1% w/w) coupled with neem leaves powders (@ 1q/ha) was most effective in managing maize cyst nematode. Kumhar *et al.* (2018a & b) reported that use of *Calotropis procera* and *Pochonia chlamydosporia* (4%) increases the maize growth and reduces maize cyst nematode (*Heterodera zaeae*). Improvement in efficacy of disease-control and crop yield was more economically and environmentally acceptable

with integration of chicken manure with NPK, *T. harzianum* or *B. thuringiensis* (Osman *et al.*, 2018). Meena *et al.* (2020) found that *Trichoderma viride* was superior in declining nematode population and per cent disease incidence along with increased growth of tomato. Similar results were also recorded with application of NSKE in tomato (Meena *et al.*, 2021b). Hot water + tobacco churi + *Paecilomyces lilacinus* combination were significantly reduced nematode reproduction and enhanced cucumber growth (Bhati *et al.*, 2021). Bhati *et al.* (2022a & b) also recorded increased cucumber yield with organic amendments and *Trichoderma viride* (5.0g per plant) along with reduced nematode reproduction.

The accumulation of peroxidase (PO), polyphenol oxidase (PPO), phenylalanine ammonia lyase (PAL) and phenol were estimated in fennel roots through spectrophotometer (Table 2). Results revealed that all the tested bio-agents and botanicals significantly enhanced the level of PO (0.788 $\mu\text{mol/min/g}$), PPO (0.094 $\mu\text{mol/min/g}$), PAL (0.646 $\mu\text{mol/min/g}$) and phenol (3.110 mg/g) activities in fennel roots over untreated control (0.423, 0.027, 0.117 $\mu\text{mol/min/g}$ and 1.513 mg/g, respectively). Maximum enzymatic activity was found in *T. harzianum* + garlic treated plants followed by *T. harzianum* + parthenium and parthenium in comparison to untreated check. The present findings are in accordance with results of Singha *et al.* (2011), Zareena *et al.* (2014), Chandrawat *et al.* (2018, 2020 a & b) and Dhayal *et al.* (2022). Singha *et al.* (2011) observed higher phenolic accumulation in *Fusarium* infected plants in the comparison of healthy. Similarly, the application of *Pseudomonas fluorescens* and *Trichoderma viride* (@ 4g/kg soil) was the best in improving plant growth, reduction in root knot nematode (*M. incognita*) reproduction and increased PO, PPO, PAL and SOD activity of tomato (Chandrawat *et al.*, 2018) as well as in chilli (Chandrawat *et al.*, 2020a). Results were also found similar with application of castor, mahua, mustard and karanj oil-cakes (@ 2.5 and 5.0 q/ha) and neem (@ 5.0 q/ha) in tomato (Chandrawat *et al.*, 2020b) in chilli (Chandrawat *et al.*, 2022) against root knot nematode, *M. incognita*. Goel and Paul (2014) evaluated that disease severity were reduced in the neem extract treated plants by inducing activities of peroxidase and lipoxygenase. Report of various workers showed enhanced plant growth and reduced nematode population as well as galling of root with oil-cakes and bio-agents combined application. Effective results were also reported with integration of neem cake and VAM in brinjal (Bora *et al.*, 2004). The integration of mustard, neem and castor oil-cakes with AMF (Bhardwaj *et al.*, 2006);

Table 1 : Management of root rot disease of fennel and nematode population through bio-agents and botanicals.

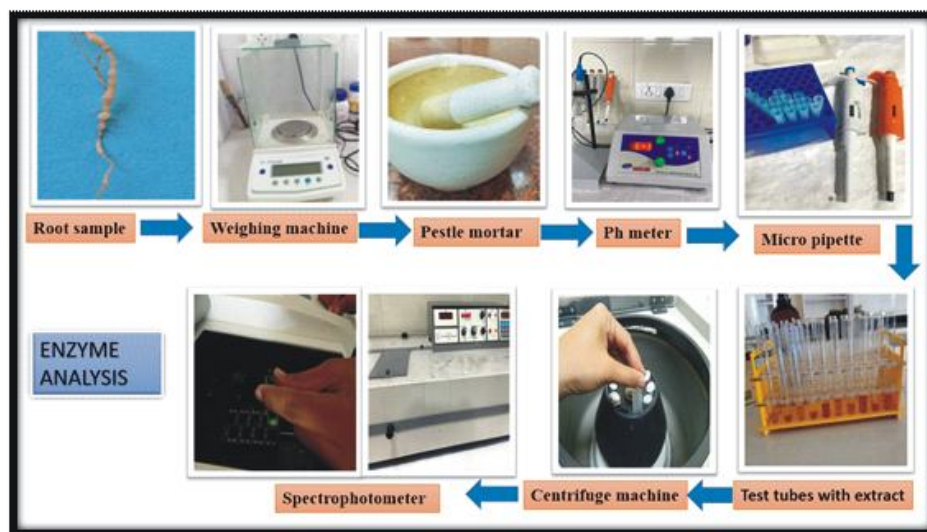
Bio-agents/ botanicals	Plant growth parameters				Root rot incidence (%) *	Nematode reproduction**			
	Shoot length (cm)	Shoot weight (g)	Root length (cm)	Root weight (g)		No. of galls/plant	No. of egg masses/Plant	No. of eggs/ egg mass	Nematode juvenile/200cc soil
<i>T. harzianum</i>	13.23	1.29	12.86	0.94	35.18(36.35)	23.34(4.98)	19.00(4.47)	284.00(16.88)	626.67(25.05)
<i>P. fluorescens</i>	8.90	0.62	6.57	0.44	67.03(54.93)	25.67(5.16)	20.34(4.61)	277.00(16.67)	764.67(27.67)
Parthenium	22.87	2.79	20.30	1.75	17.40(24.64)	13.34(3.78)	10.34(3.36)	255.00(16.00)	329.67(18.18)
Garlic	19.84	2.25	17.54	1.47	22.59(28.36)	18.34(4.39)	14.34(3.91)	269.34(16.44)	525.34(22.94)
<i>T. harzianum</i> + parthenium	15.80	1.87	15.27	1.25	14.81(22.60)	10.67(3.41)	8.67(3.10)	248.67(15.80)	263.34(16.25)
<i>T. harzianum</i> + garlic	25.50	3.51	21.94	2.35	11.48(19.76)	6.67(2.76)	3.67(2.15)	238.67(15.48)	163.67(12.83)
<i>P. fluorescens</i> + parthenium	10.37	0.88	8.24	0.66	19.25(26.01)	15.00(3.99)	10.67(3.41)	262(16.21)	424.34(20.62)
<i>P. fluorescens</i> + garlic	11.90	0.91	10.54	0.87	25.55(30.34)	21.67(4.76)	18.67(4.43)	274.34(16.59)	586.67(24.24)
Control	6.24	0.43	4.70	0.37	71.11(57.46)	29.34(5.50)	23.34(4.93)	298.67(17.31)	874.34(29.58)
SEm±	0.19	0.02	0.20	0.01	0.63	0.08	0.09	0.02	0.03
CD 5%	0.59	0.06	0.61	0.05	1.91	0.24	0.28	0.07	0.10

Average of three replications, PDI = Per cent disease incidence, *Figures given in parentheses are angular transformed, ** Figures given in parentheses are square root $\left(\sqrt{x+0.5}\right)$ transformed, Bio-agents = Soil application (@ 2.5kg/ha), Botanicals = Seed soaking (@ 10% for 30 min).

Table 2 : Effect of phyto-extracts and bio-agents on PO, PPO, PAL and Phenol concentration in fennel roots.

Treatments	Concentration of enzymes and phenol			
	PO ($\mu\text{mol/min/g}$)	PPO ($\mu\text{mol/min/g}$)	PAL ($\mu\text{mol/min/g}$)	Phenol (mg/g)
<i>T. harzianum</i>	0.621	0.042	0.212	1.910
<i>P. fluorescens</i>	0.593	0.040	0.182	1.826
Parthenium	0.677	0.076	0.442	2.080
Garlic	0.648	0.053	0.382	2.043
<i>T. harzianum</i> + parthenium	0.734	0.080	0.554	2.423
<i>T. harzianum</i> + garlic	0.788	0.094	0.646	3.110
<i>P. fluorescens</i> + parthenium	0.675	0.073	0.427	2.050
<i>P. fluorescens</i> + garlic	0.646	0.043	0.377	2.030
Control	0.423	0.027	0.117	1.513
SEm\pm	0.001	0.000	0.000	0.009
CD 5%	0.003	0.000	0.001	0.029

Average of three replications, PDI= Per cent disease incidence, *Figures given in parentheses are angular transformed, ** Figures given in parentheses are square root () transformed, PO- Peroxidase, PPO- Polyphenol oxidase, PAL- Phenylalanine ammonia lyase, Bio-agents- Soil application (@ 2.5kg/ha), Botanicals- Seed soaking (@ 10% for 30 min)

**Fig. 1 :** Enzyme analysis process.

neem cake with *Pasteuria penetrans* (Javed *et al.*, 2008) in tomato and integration of *Pupurescillium lilacinus* with groundnut cake in eggplant (Ashraf *et al.*, 2010) and castor cake and *P. penetrans* combination in chilli (Chaudhary *et al.*, 2013) proved effective. Combination of neem cake, mustard cake, neem leaf extract and castor extract with *P. lilacinus* and *T. harzianum* against root nematode of brinjal and found most effective in managing the disease (Zareena *et al.*, 2014). Dhayal *et al.* (2022) observed that neem extract @ 10 % concentration was found best to enhance PPO, PAL and phenol activity in tomato. Choudhary *et al.* (2023) found that neem cake (10%) was superior to enhance PPO, PAL and phenol activity.

Conclusion

The use of bio-agents and botanicals is beneficial in reducing disease incidence and also in increasing the level of defense enzymes in plants as compared to diseased plants which also plays important role in reduced pathogen population and enhanced plant growth parameters. Soil application of *Trichoderma harzianum* (@ 2.5kg/ha) + seed soaking in garlic extract (10% for 30 min) was proved effective in enhancing plant biomass and in reducing the disease incidence. In conclusion, the use of the *T. harzianum* + garlic extract can be considered as a tool complementary for the commercial management of root rot of fennel in areas where this disease is gaining importance and becoming a limiting factor. Additionally, it can be used for managing pathogen resistance as well

as also provide opportunity for organic production of spices for consumers.

Competing interests : Authors have declared that no competing interest exist.

Authors' contributions

Methodology: 'Kiran Kumawat' conducted the original experiment, 'S. K. Goyal' and 'B. S. Chandrawat' wrote original draft manuscript, 'G. K. Mittal' and 'R. P. Ghasolia' done biochemical and statistical analysis, 'Lalita Lakhran' and 'B. L. Naga' reviewed and edited the MS and 'Dinesh Kumar Meena and Sushila Yadav' managed the literature.

References

- Akhtar, H., Arif A. and Anita S. (2007). Disease complex in *Pisum sativum* involving *Meloidogyne incognita* and *Fusarium oxysporum* f.sp. *pisi*. *Annals Plant Protect. Sci.*, **15**(1), 189-194.
- Anonymous (2020-21). Rajasthan Agricultural Statistics at a glance.
- Arya, R. (2016). Combined effect of *Trichoderma harzianum* and vermicompost in the management of root-knot nematode (*Meloidogyne incognita*) in tomato cv Pusa Ruby. *Indian Phytopathology*, **69**, 464-467.
- Ashraf, M.S. and Khan T.A. (2010). Integrated approach for the management of *Meloidogyne javanica* on eggplant using oil cakes and biocontrol agents. *Arch. Phytopathol. Plant Protect.*, **43**, 609-614.
- Atkinson, G.F. (1892). Some diseases of cotton. *Alabama Agricult. Exp. Station Bull.*, **41**, 65.
- Baheti, B.L., Meena K.K., Chandrawat B.S., Khandelwal S.K. and Singh D. (2016). Botanicals: An effective Tool to Check the Initial Infection of Maize Cyst Nematode, *Heterodera zae* on Maize as Seed treatment. *Adv. Life Sci.*, **5**(11), 4502-4506.
- Bhagawati, B. and Goswami B.K. (2000). Interaction of *Meloidogyne incognita* and *Fusarium oxysporum* f. sp. *lycopersici* on tomato. *Indian J. Nematol.*, **30**(1), 93-94.
- Bhardwaj, A. and Sharma S. (2006). Biocontrol of *Meloidogyne incognita* in *Lycopersicon esculentum* with AM fungi and oil cakes. *Plant Pathol. J.*, **5**, 166-172.
- Bhati, S.S., Baheti B.L., Chandrawat B.S. and Ahuja A. (2022a). Utilization of agri-based wastes against *Meloidogyne incognita* infecting cucumber in polyhouse. *Indian J. Nematol.*, **52**(2), 155-163.
- Bhati, S.S., Baheti B.L. and Chandrawat B.S. (2021). Influence of Eco-Friendly Management Modules for Suppression of *Meloidogyne incognita* infecting Cucumber in Polyhouse. *Indian J. Nematol.*, **51**(1), 27-33.
- Bhati, S.S., Baheti B.L., Singh I. and Chandrawat B.S. (2022b). Impact of Bio-agents against *Meloidogyne incognita* infecting Cucumber in Poly- House. *Biological Forum – An Int. J.*, **14**(2), 1481-1487.
- Bora, A. and Phukan P.N. (2004). Comparative efficacy of *Glomus fasciculatum* with neem cake and carbofuran for the management of *Meloidogyne incognita* on brinjal. *Indian J. Nematol.*, **34**, 129-132.
- Chandrawat, B.S., Siddiqui A.U., Bhati S.S. and Saharan V. (2018). Induction of defense enzymes using bio-agents in tomato infected with root knot nematode, *Meloidogyne incognita*. *Indian J. Nematol.*, **48**(2), 139-145.
- Chandrawat, B.S., Siddiqui A.U., Bhati S.S. and Saharan V. (2020a). Bio-agents: a source for initiation of defense enzymes in chilli infected with root-knot nematode, *Meloidogyne incognita*. *J. Entomol. Zool. Stud.*, **8**(6), 1684-1688.
- Chandrawat, B.S., Siddiqui A.U. and Saharan V. (2022). Determination of defense enzyme induction in chilli in response to root- knot nematode, *Meloidogyne incognita* upon oil cake application. *Indian J. Nematol.*, **52**(2), 225-233.
- Chandrawat, B.S., Siddiqui A.U., Bhati S.S. and Saharan V. (2020b). Response of defense related enzymes in tomato treated with oil-cakes against root-knot nematode, *Meloidogyne incognita*. *Int. J. Curr. Microbiol. Appl. Sci.*, **9**(11), 1100-1111.
- Chaudhary, K.K. and Kaul R.K. (2013). Efficacy of *Pasteuria penetrans* and various oil seed cakes in management of *Meloidogyne incognita* in chilli pepper (*Capsicum annuum* L.). *J. Agricult. Sci. Technol.*, **15**, 617-626.
- Choudhary, K., Bishnoi S.P., Dhayal R., Chandrawat B.S., Mittal G.K. and Gurjar H. (2023). Defense response in brinjal plants treated with oil-cakes against root-knot nematode, *Meloidogyne javanica*. *Agricult. Mechanization in Asia, Africa and Latin America*, **54**(4), 12875-12887.
- Dhayal, R., Chandrawat B.S., Choudhary K., Mittal G.K., Gurjar H., Bishnoi S.P. and Sharma R.L. (2022). Response of Botanicals and Enzymes Activities as a Defense Mechanism against Root-knot nematode, *Meloidogyne javanica* in Tomato. *Agricultural Mechanization in Asia*, **53**(9), 9753-9766.
- Dickerson, D.P., Pascholati S., Hagerman A.E., Butler L.G. and Nicholson R.L. (1984). Phenylalanine ammonia lyase and hydroxycinnamate: CoA ligase in maize mesocotyls inoculated with *Helminthosporium maydis* or *H. carbonzlm*. *Physiological Plant Pathology*, **25**, 111-123.
- Goel, N. and Paul P.K. (2014). Neem fruit extract induces peroxidase and lipoxygenase in tomato. *Asian J. Biolog. Life Sci.*, **3**, 189-194.
- Gupta, J.H. and Srivastava V.P. (1976). A new root rot of fennel caused by *Fusarium solani*. *J. Mycol. Plant Pathol.*, **8**, 206.
- Hammerschmidt, R., Nuckles E.M. and Kuc J. (1982). Association of enhanced peroxidase activity with induced systemic resistance of cucumber to *Colletotrichum lagenarium*. *Physiological Plant Pathology*, **20**, 73-82.

- Javed, N., El-Hassan S., Gowen S., Pemproke B. and Inam-ul-Haq M. (2008). The potential of combining *Pasteuria penetrans* and neem (*Azadirachta indica*) formulations as a management system for root-knot nematodes on tomato. *Europ. J. Plant Pathol.*, **120**, 53-60.
- Kamal, A.M.A.E., Magd E.M.A. and Abdel M.A. (2009). Management of tomato root-knot nematode *Meloidogyne incognita* by plant extracts and essential oils. *The Plant Pathol. J.*, **25**(2), 189-192.
- Kant, K., Meena S.R. and Sharma Y.K. (2013). Root-knot infestation in cumin crop. *Seed Spices-E-Newsletter*, **5**(5), 2.
- Khare, M.N., Tiwari S.P. and Sharma Y.K. (2014). Disease problems in fennel (*Foeniculum vulgare* Mill) and fenugreek (*Trigonella foenum graceum* L.) cultivation and their management for production of quality pathogen free seeds. *Int. J. Seed Spices*, **4**(2), 11-17.
- Kumar, D., Bhatt J., Sharma R.L. and Kumar N. (2017). Interaction between *Meloidogyne incognita* and *Fusarium oxysporum* on Black gram (*Vigna mungo* L.). *Int. J. Chem. Stud.*, **5**(4), 624-627.
- Kumawat, K., Goyal S.K., Chandrawat B.S., Mittal G.K., Ghasolia R.P., Teli M., Kansotia K. and Sharma P. (2022). Interactive effect of *Meloidogyne javanica* and *Fusarium solani* in causing root rot of fennel. *Annals Plant Protect. Sci.*, **30**(2), 86-90.
- Kumhar, R.N., Baheti B.L. and Chandrawat B.S. (2018a). Eco-friendly management of maize cyst nematode, *Heterodera zae* on maize by use of botanicals. *Int. J. Curr. Microbiol. Appl. Sci.*, **7**(6), 199-204.
- Kumhar, R.N., Baheti B.L., Chandrawat B.S. and Gurjar O.P. (2018b). Seed treatment with bio-agents: An environmental suitable approach for management of maize cyst nematode, *Heterodera zae* on maize. *Curr. Nematol.*, **29**(1,2), 1-5.
- Mai, W.F. and Abawi G.S. (1987). Interactions among root knot nematodes and *Fusarium* wilt fungi on host plants. *Annu. Rev. Phytopathol.*, **25**, 317-338.
- Malick, C.P. and Singh M.B. (1980). Plant enzymology and histo-enzymology. Kalyani publications, New Delhi, 286.
- Mayer, A.M., Harel E. and Shaul R.B. (1965). Assay of catechol oxidase a critical comparison of methods. *Phytochemistry*, **5**, 783-789.
- McBeth, C.W., Taylor A.L. and Smith A.L. (1941). Note on staining nematodes in root tissues. *Proceeding of Helminthological Society of Washington*, **8**, 26.
- Meena, K.K., Baheti B.L., Rathore B.S. and Chandrawat B.S. (2015). Organic amendment-an important tool for the management of maize cyst nematode, *Heterodera zae* on maize (*Zea mays* L.). *Curr. Nematol.*, **26**(1,2), 23-27.
- Meena, K.K., Baheti B.L., Rathore B.S. and Chandrawat B.S. (2016). Integration of chemical and botanicals for the management of maize cyst nematode, *Heterodera zae* on maize. *Ecol. Environ. Conser.*, **22**, 151-154.
- Meena, M., Chandrawat B.S., Ahir R.R., Dhayal R., Choudhary A., Kavita and Meena A.K. (2023). Effect of *Fusarium oxysporum* and *Meloidogyne* spp. interaction on wilt disease incidence in cucumber. *Annals Plant Protect. Sci.*, **31**(1), 39-43.
- Meena, P., Chandrawat B.S. and Ahir R.R. (2020). Bioagents for management of wilt complex in tomato by *Meloidogyne incognita* and *Fusarium oxysporum* f. sp. *lycopersici*. *Annals Plant Protect. Sci.*, **28**(2), 160-162.
- Meena, P., Chandrawat B.S., Ahir R.R., Maurya S., Lakhra L., Prajapati S. and Kumawat C.K. (2021a). Effect of *Fusarium oxysporum* f. sp. *lycopersici* and *Meloidogyne incognita* interaction on wilt disease incidence in tomato. *Int. J. Chem. Stud.*, **9**(1), 2621-2624.
- Meena, P., Chandrawat B.S., Ahir R.R., Meena A.K. and Singh M. (2021b). Management of tomato root-knot wilt complex caused by *Meloidogyne incognita* and *Fusarium oxysporum* f. sp. *lycopersici* through plant-extracts. *Int. J. Chem. Stud.*, **9**(1), 3100-3103.
- Omidbaigi, R. (2000). Production and Processing of Medicinal Plants. Astan Quds Publication, Tehran. *Acta Universitatis Sapientiae Agriculture and Environment*, **3**, 132-143.
- Osman, H.A., Ameen H.H., Mohamed M., El-Mohamedy A. and Elkelany U.S. (2018). Field control of *Meloidogyne incognita* and root rot disease infecting eggplant using nematicide, fertilizers and microbial agents. *Egypt. J. Biolog. Pest Control*, **28**(40), 44-1.
- Park, J., Seo Y. and Kim Y.H. (2014). Biological control of *Meloidogyne hapla* using an antagonistic bacterium. *The Plant Pathol. J.*, **30**(3), 288-298.
- Patel, A.D. (2011). Nematode diseases of seed spices and their management. *Recent Advances in Seed Spices*, **163**, 5.
- Patel, R.G., Patel H.R., Vyas R.V., Patel B.A. and Patel B.N. (2002). Assessment of avoidable yield losses due to root-knot nematode, *Meloidogyne javanica* pathotype 2 in fennel field. *Proceedings of National Symposium on Biodiversity and Management of Nematodes in Cropping Systems for Sustainable Agriculture*, Jaipur, India. 131-132.
- Singha, M.I., Kakoty Y., Unni B.G., Kalita M.C., Das J., Naglot A., Wann S.B. and Singh L. (2011). Control of *Fusarium* wilt of tomato caused by *Fusarium oxysporum* f.sp. *lycopersici* using leaf extract of *Piper betel* L. *World J. Microbiol. Biotechnol.*, **1730**, 6.
- Taylor, A.L. and Sasser J.N. (1978). Biology, identification and control of root knot nematode (*Meloidogyne* spp.). North California State University graphs, Raleigh, N.C., 111.
- Taylor, D.P. and Netscher G. (1974). An improved technique of preparing perineal patterns of *Meloidogyne* spp. *Nematologica*, **20**, 268- 269.
- Zareena, S.K. and Das V. (2014). Vanita. Root-knot disease and its management in brinjal. *Glob. J. Bio-Sci. Biotechnol.*, **3**(1), 126-127.